

CSF Phosphoproteomics Biomarker Analysis from the Phase 2 Clinical Trial SHINE to Elucidate the Role of CT1812 in Alzheimer's Disease

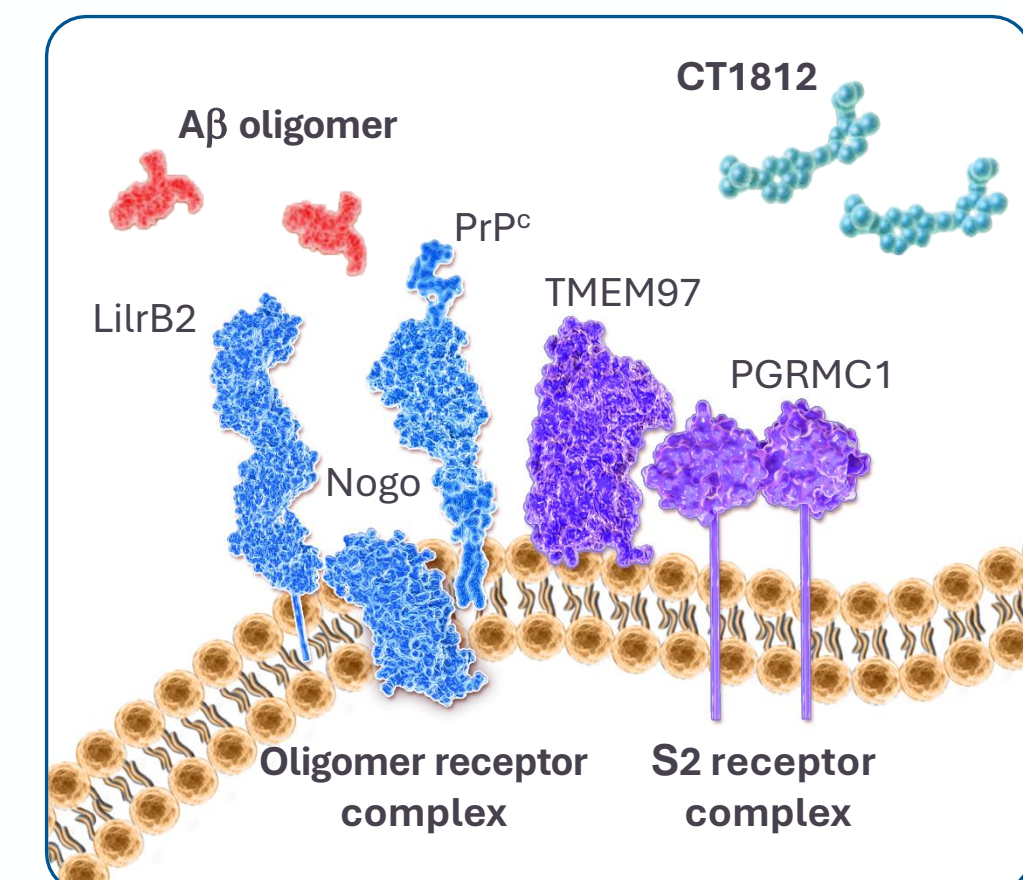
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Key takeaway: For the first time we showed that the S2R modulator, CT1812 impacts phosphorylation of AD-related proteins (e.g. APOE, CHGB) in CSF samples from Phase 2 clinical trials for Alzheimer's disease.

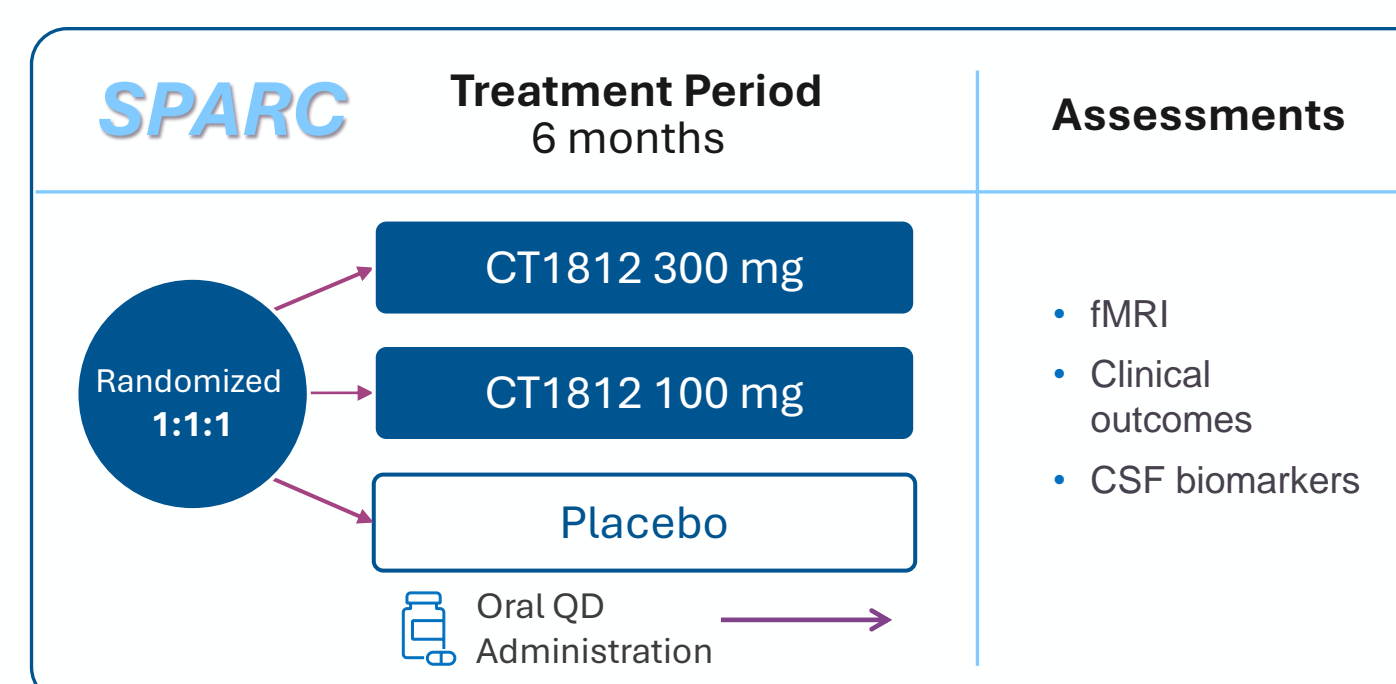
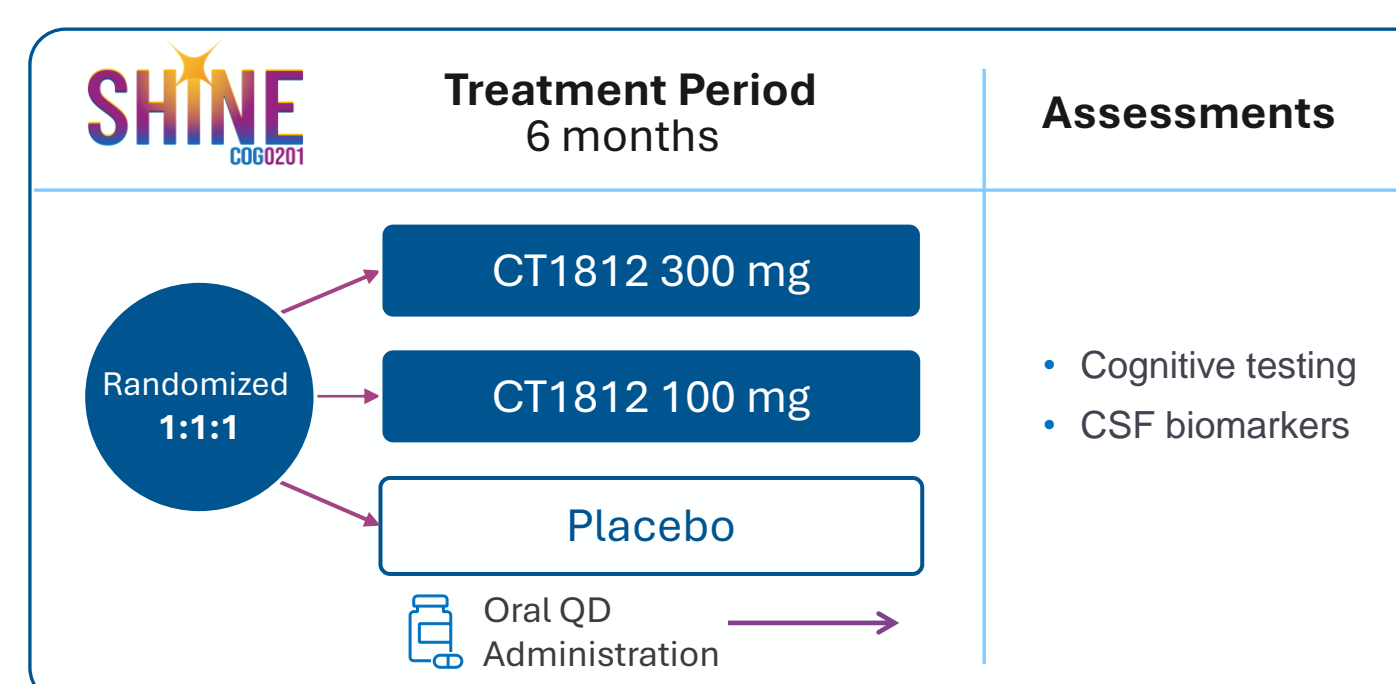
Introduction

The sigma-2 receptor (S2R) modulator, CT1812, is allosteric Aβ oligomer antagonist currently in Phase 2 clinical trials¹ for Alzheimer's disease (AD). Preclinical and clinical studies have shown that CT1812 displaces Aβ oligomers from synapses² which may facilitate clearance of Aβ oligomers in the cerebrospinal fluid, restoring cognitive performance in a transgenic mouse model of AD³.



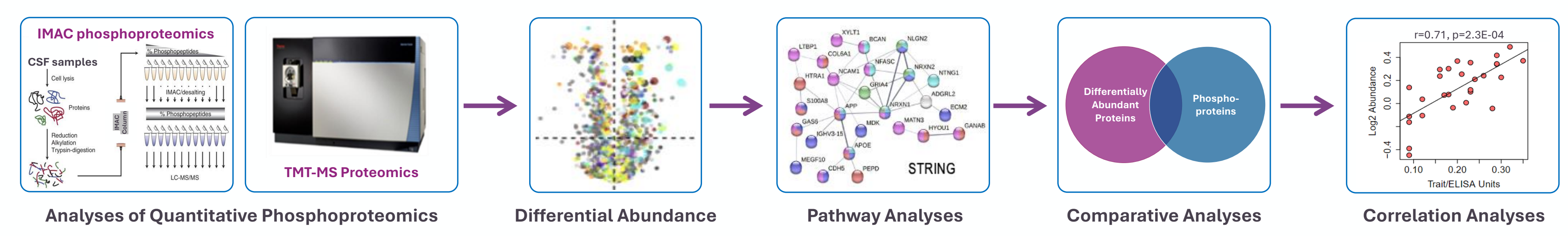
To investigate the mechanism of action of CT1812 and enable biomarker discovery, a phosphoproteomic analysis of CSF samples from SHINE-A was performed. SHINE is a Phase 2 randomized, double-blind, placebo-controlled trial to assess the safety and tolerability of two doses of CT1812 for 6 months in mild-to-moderate AD patients (NCT03507790), and an interim analysis was performed on the first 24 patients enrolled (SHINE-A).

Methods



- Tandem-mass tag mass spectrometry (TMT-MS) proteomics and immobilized metal affinity chromatography (IMAC) phosphoproteomics were performed on baseline and end-of-study CSF samples from SHINE-A (N=18) and differential abundance (CT1812 vs placebo; $p \leq 0.05$) was assessed.
- STRING pathway analyses (v12.0) were conducted on total phosphoproteins.
- Pearson correlation analyses with altered phosphopeptides and CSF Aβ42 level and ADAS-Cog-11 score were performed.
- Clinical validation of findings were conducted on an independent cohort, SPARC (NCT03493282, N=23). The study populations were similar and reference Fig. 5D below.

Workflow



Results

CT1812 Treatment Impact on CSF Phosphoproteome

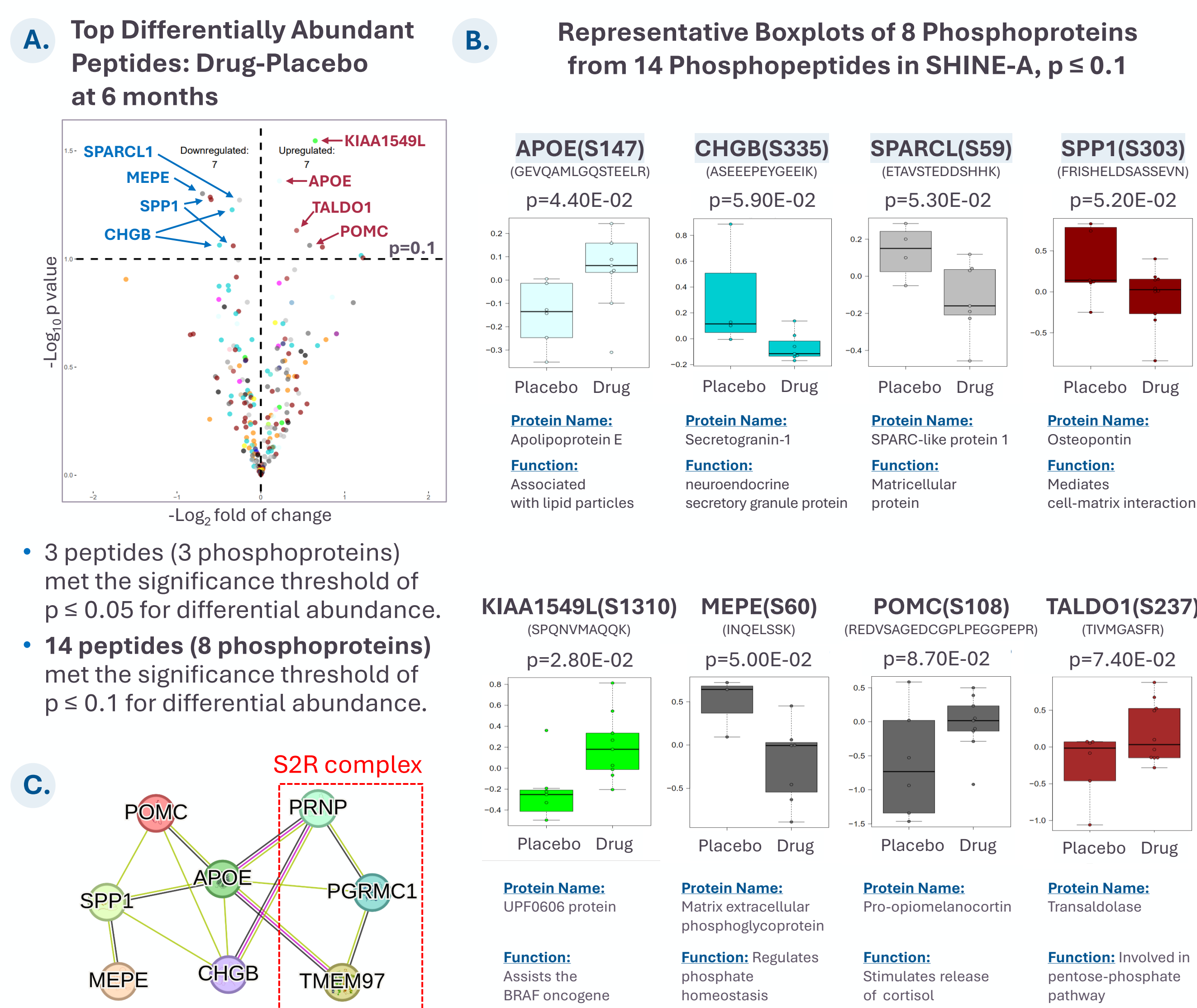


Figure 1. (A) Volcano plots to visualize the global phosphoproteomics change between CT1812-treated and placebo samples. Each data point in the scatter plot represents a protein, $p \leq 0.1$. **(B)** Identified 8 phosphoproteins, from 14 phosphopeptides, altered in SHINE-A CSF samples. Representative boxplots of the identified phosphopeptides involved AD-related proteins highlighted in blue ($p \leq 0.1$). **(C)** Protein-protein interaction map with S2R complex components via STRING (v12.0). TMEM97 (S2R), PRNP, and PGRMC1 were added to understand how they interacted with the identified proteins (list in B).

Pathways Related to Aβ and Lipoprotein are Impacted by CT1812 in CSF of Treated-Patients

GO terms	Term description	Strength	p-value
1902998	Positive regulation of neurofibrillary tangle assembly	2.34	2.08E-02
1905908	Positive regulation of amyloid fibril formation	2.21	1.70E-03
0032805	Positive regulation of LDL particle receptor catabolic process	2.16	2.81E-02
1902947	Regulation of tau-protein kinase activity	1.82	8.80E-04
1900221	Regulation of amyloid-beta clearance	1.53	2.15E-02

N.B. Top 5 most relevant GO terms were selected.

Figure 2. STRING pathway analysis (v12.0) of phosphoproteomics data with total 206 phosphopeptides (91 phosphoproteins). GO=gene ontology.

CT1812 Treatment Altered Proteins' Phosphorylation and Abundance

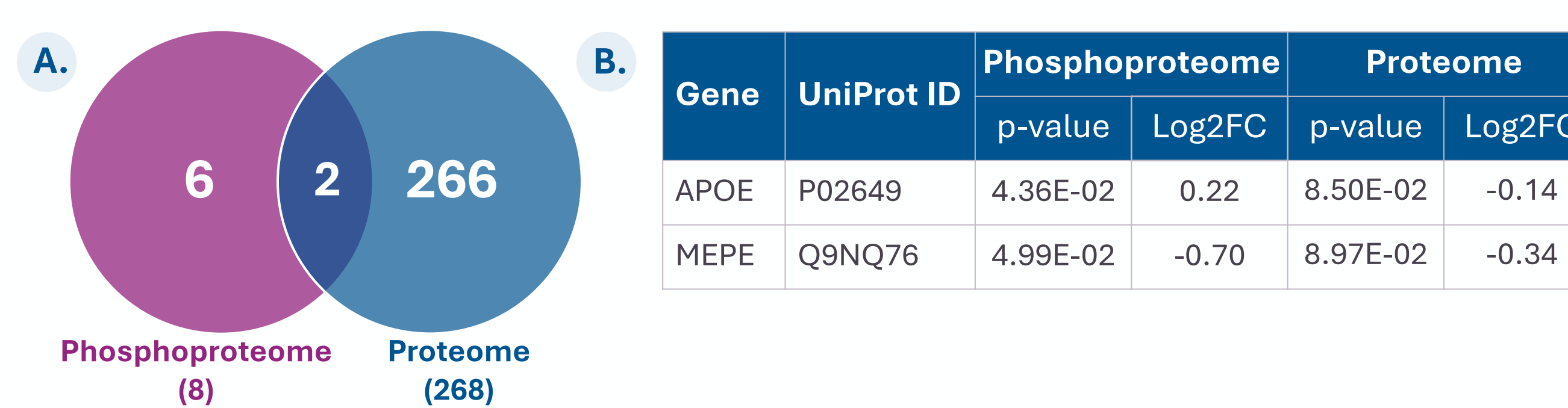


Figure 3. (A) Venn diagram to show overlapping proteins between SHINE-A phosphoproteins and differentially abundant proteins ($p \leq 0.1$). **(B)** p-value and fold change (Log2FC) for the 2 common proteins.

Correlated Phosphopeptides with CSF Aβ42 Levels and ADAS-Cog-11 Score were Identified

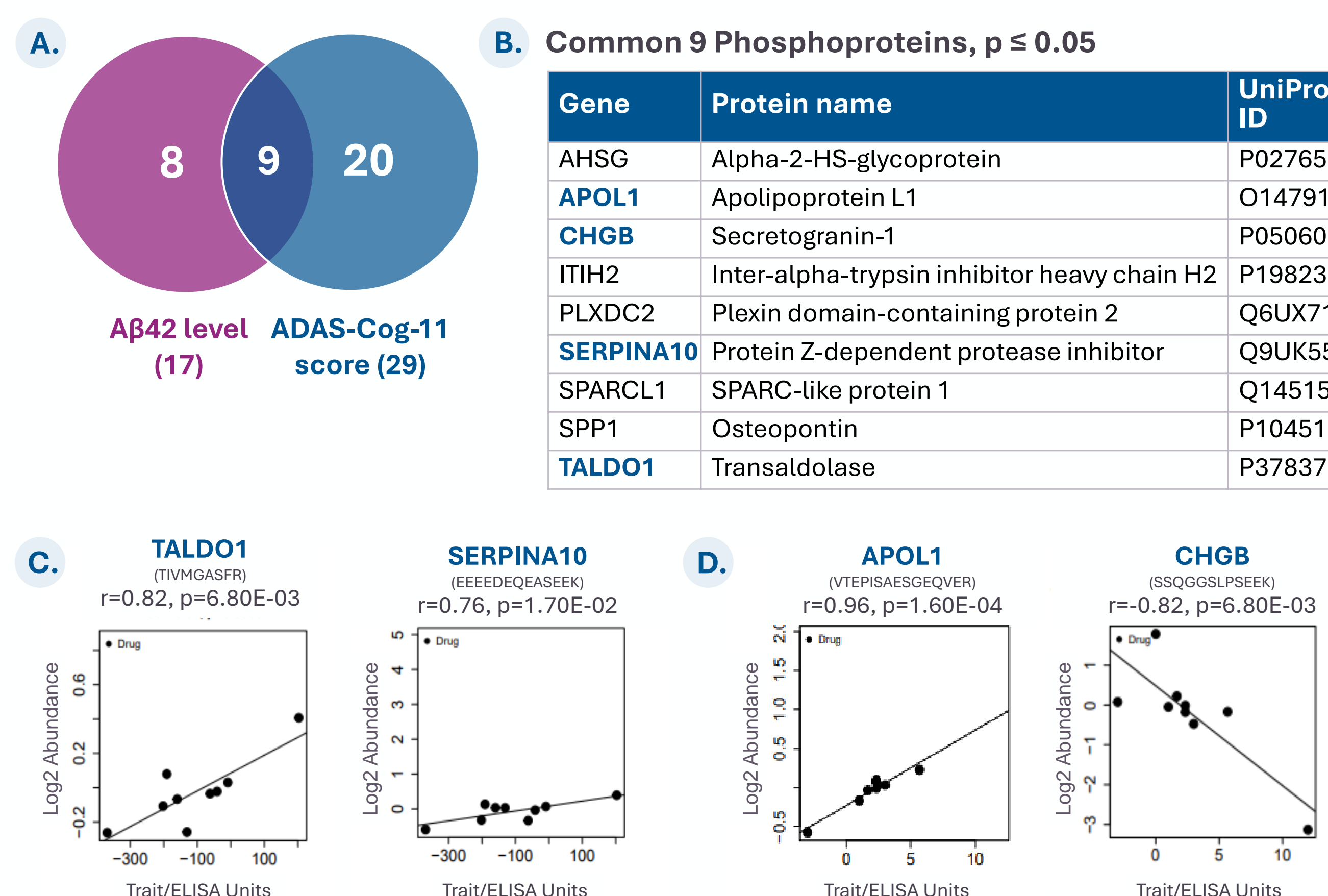


Figure 4. Comparative analysis between phosphopeptides correlates with CSF Aβ42 levels and ADAS-Cog-11 score in CT1812-treated group only by Pearson's correlation analysis. **(A)** Venn diagram to show common correlates ($p \leq 0.05$). **(B)** List of correlated phosphoproteins. **(C)** and **(D)** Representative scatter plots of correlated phosphopeptides with Aβ42 level and ADAS-Cog-11 score, respectively ($p \leq 0.05$, $r \pm [0.75]$).

Conclusions

- The S2R modulator CT1812 mediates AD-related proteins' phosphorylation in CSF samples from Phase 2 clinical trials for Alzheimer's disease.
- Pathways enriched by CT1812 in SHINE-A are related to vesicle trafficking, neurofibrillary tangle assembly, amyloid fibril formation, low-density lipoprotein particle receptor catabolic processes, and tau-protein kinase activity.
- Identified phosphopeptides were correlated with Aβ42 levels and ADAS-Cog-11 score in participants treated with CT1812 in SHINE-A.
- Altered phosphoproteins by S2R modulator CT1812 in SHINE-A replicated in an independent cohort, SPARC.

Major AD-Related Pathways and Four Phosphoproteins Replicated in SPARC, an Independent Cohort

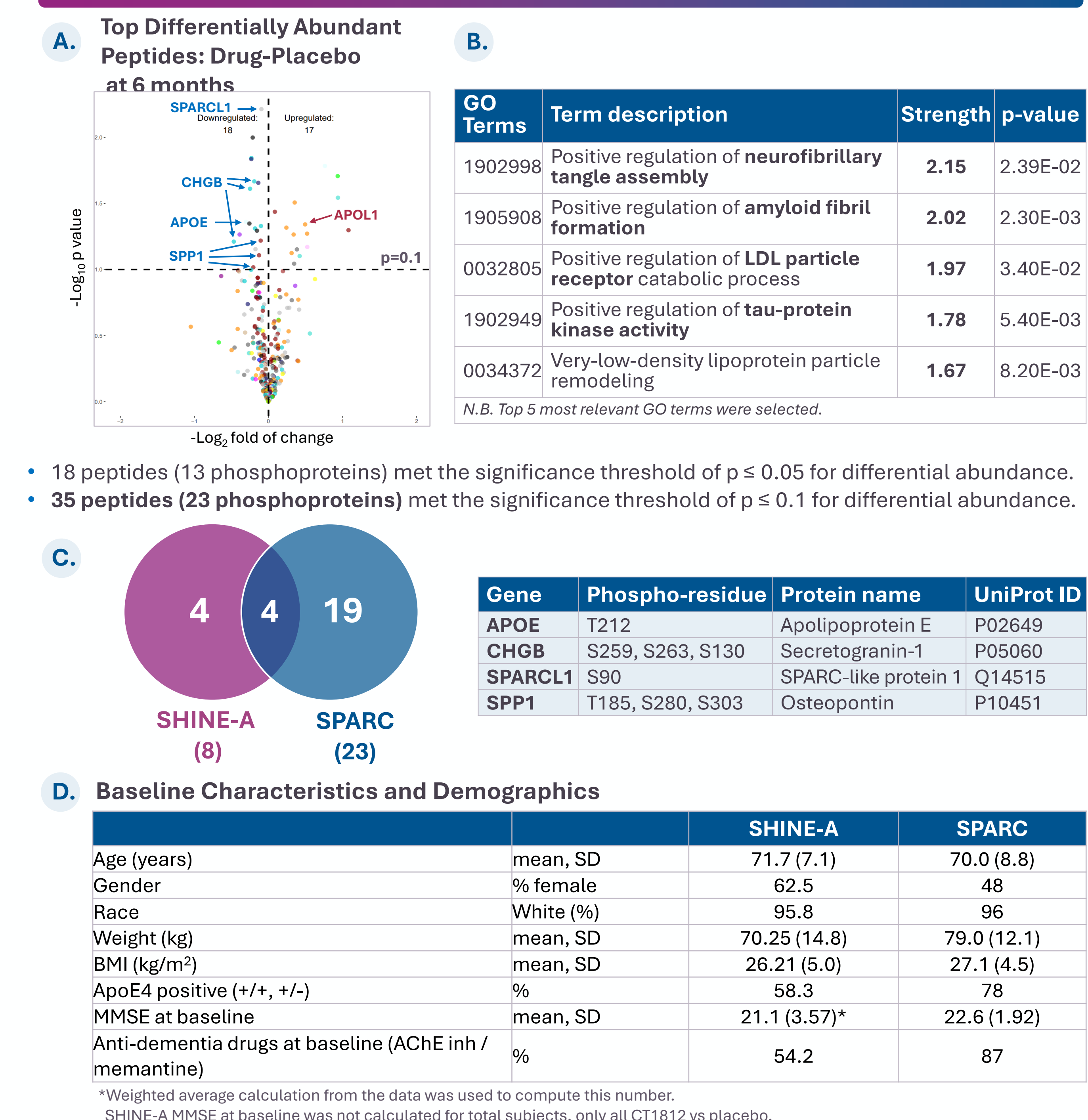


Figure 5. (A) Volcano plots to visualize the global phosphoproteomics change between CT1812-treated and placebo samples in the SPARC trial. Each data point in the scatter plot represents a protein, $p \leq 0.1$. **(B)** STRING pathway analysis (v12.0) of phosphoproteomics data with total 303 phosphopeptides (140 phosphoproteins). GO=gene ontology. **(C)** Venn diagram to show overlapping phosphorylated proteins by CT1812 between SHINE-A and SPARC trials, and the list of common phosphoproteins ($p \leq 0.1$). AD-related proteins highlighted in blue. **(D)** Baseline Characteristics and Demographics to show comparability between cohorts.

